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(54) Title: HER2 EXTRACELLULAR DOMAIN (57) Abstract An extracellular portion of the HER2 molecule, essentially free of transmembrane and cytoplasmic portions, which is antigenic in animals. Isolated DNA encoding the extracellular portion; an expression vector containing the isolated DNA; and a cell containing the expression vector. A process for producing the extracellular domain. A vaccine containing the extracellular domain.		

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-1-

HER2 EXTRACELLULAR DOMAINBACKGROUND OF THE INVENTIONField of the Invention

The present invention is generally directed to the
5 extracellular domain of p185^{HER2}, a receptor-like protein
which is encoded by the human homolog of the rat neu
oncogene.

More specifically, the present invention is directed
to a form of the extracellular domain which is essentially
10 free of transmembrane and cytoplasmic domains, to the DNA
encoding this form, and to a process for producing this
form of the extracellular domain in a host cell.

Description of Background and Relevant Materials

Human epidermal growth factor receptor 2 (HER2, also
15 known as NGL and human c-erbB-2, or ERBB2), is the human
homolog of the rat proto-oncogene neu. HER2 encodes a
1,255 amino acid tyrosine kinase receptor-like glycoprotein
with homology to the human epidermal growth factor
receptor. Although no ligand binding to this probable
20 growth factor receptor has yet been isolated, the HER2 gene
product, p185^{HER2}, has the structural and functional
properties of subclass I growth factor receptors (Yarden et al.,
Ann. Rev. Biochem., 57:443-478 (1988); Yarden et al.,
Biochem., 27:3113-3119 (1988)).

25 The receptor tyrosine kinases all have the same
general structural motif; an extracellular domain that
binds ligand, and an intracellular tyrosine kinase domain
that is necessary for signal transduction, or in aberrant

-2-

cases, for transformation. These 2 domains are connected by a single stretch of approximately 20 mostly hydrophobic amino acids, called the transmembrane spanning sequence. This sequence is thought to play a role in transferring the
5 signal generated by ligand binding from the outside of the cell to the inside. It has also been suggested to play a role in the proper positioning of the receptor in the plasma membrane.

Consistent with this general structure, the p185^{HER2}
10 glycoprotein, which is located on the cell surface, may be divided into three principle portions: an extracellular domain, or ECD (also known as XCD); a transmembrane spanning sequence; and a cytoplasmic, intracellular tyrosine kinase domain. While it is presumed that the
15 extracellular domain is a ligand receptor, as stated above the corresponding ligand has not yet been identified.

The HER2 gene is of particular interest because its amplification has been correlated with certain types of cancer. Amplification of the HER2 gene has been found in
20 human salivary gland and gastric tumor-derived cell lines, gastric and colon adenocarcinomas, and mammary gland adenocarcinomas. Semba et al., Proc. Natl. Acad. Sci. USA, 82:6497-6501 (1985); Yokota et al., Oncogene, 2:283-287 (1988); Zhou et al., Cancer Res., 47:6123-6125 (1987); King
25 et al., Science, 229:974-976 (1985); Kraus et al., EMBO J., 6:605-610 (1987); van de Vijver et al., Mol. Cell. Biol., 7:2019-2023 (1987); Yamamoto et al., Nature, 319:230-234 (1986). Gene transfer experiments have shown that

-3-

overexpression of HER2 will transform NIH 3T3 cells and also cause an increase in resistance to the toxic macrophage cytokine tumor necrosis factor. Hudziak *et al.*, "Amplified Expression of the HER2/ERBB2 Oncogene Induces
5 Resistance to Tumor Necrosis Factor Alpha in NIH 3T3 Cells", Proc. Natl. Acad. Sci. USA 85, 5102-5106 (1988).

Because amplification of the HER2 gene results in greatly increased numbers of the p185^{HER2} protein residing on the surfaces of affected cells, there may be an
10 interrelationship between increased amounts of p185^{HER2} extracellular domain on the surfaces of affected cells and the resistance of these cells to treatment. It would therefore be highly desirable to be able to manipulate the p185^{HER2} extracellular domain in order to investigate several
15 possibilities for the treatment of conditions associated with amplification of the HER2 gene. These therapeutic modes relate not only to the extracellular domain, but also to the putative ligand, which it should be possible to isolate and characterize using the purified p185^{HER2}
20 extracellular domain.

SUMMARY OF THE INVENTION

The present invention is accordingly directed to an extracellular portion of the HER2 molecule containing at least 9 amino acids, and/or containing an immune epitope,
25 which is essentially free of transmembrane and intracellular portions of the HER2 molecule. The extracellular portion may be substantially pure, or at least about 99% pure, and may extend to the entire

-4-

extracellular portion of the HER2 molecule. Moreover, the extracellular portion may be antigenic in animals, and may be conjugated with a peptide having immunogenic properties; this peptide may contain an immune epitope.

5 In another embodiment, the present invention is directed to isolated DNA encoding the extracellular portion of the HER2 molecule. This isolated DNA terminates upstream of the DNA portion encoding the transmembrane domain of the HER2 molecule. The termination may occur at
10 least 1 base pair upstream of the portion encoding the transmembrane domain of the HER2 molecule, and preferably occurs about 24 base pairs upstream of this portion.

 The isolated DNA of the present invention encodes a sequence of at least 9 amino acids of the extracellular
15 portion, and none of the transmembrane or intracellular portions of the HER2 molecule.

 In a further embodiment, the present invention contemplates an expression vector, such as a plasmid or virus, containing the isolated DNA as described above, as
20 well as a cell containing the expression vector. This cell may be eukaryotic or prokaryotic.

 The present invention also extends to a process for producing an extracellular portion of the HER2 molecule, which includes the steps of ligating the isolated DNA as
25 described above into an expression vector capable of expressing the isolated DNA in a suitable host; transforming the host with the expression vector;

-5-

culturing the host under conditions suitable for expression of the isolated DNA and production of the extracellular portion; and isolating the extracellular portion from the host. The host cell may be a prokaryote, such as a
5 bacterium, or a eukaryote.

In a yet further embodiment, the present invention extends to a vaccine comprising the extracellular portion of the HER2 molecule, which may be combined with suitable adjuvants.

10

BRIEF DESCRIPTION OF FIGURES

Fig 1. HER2 expression vector and full-length and mutant HER2 proteins. The HER2 expression vector contained eukaryotic transcriptional units for the mouse dihydrofolate reductase (DHFR) cDNA and the bacterial neomycin
15 phosphotransferase (neo) gene, both under SV40 early promoter control. Transcription of the full-length HER2 cDNA was also driven by the early SV40 promoter. The full-length HER2 protein contains an extracellular domain with two cysteine-rich clusters (hatched rectangle),
20 separated by the transmembrane-spanning region (filled rectangle) from the intracellular tyrosine kinase domain (open rectangle). The mutant protein p185^{HER2ΔTM} has a deletion of 28 amino acids, including the transmembrane-spanning region. The truncated p185^{HER2XCD}
25 protein contains all N-terminal sequences up to 8 amino acids before the transmembrane-spanning region.

Fig. 2. Amplification of HER2 and HER2ΔTM genes. Cell lines transfected with plasmids expressing wild type

-6-

or the ATM mutant HER2 cDNAs were amplified to resistance to 400 nM methotrexate. Cultures were metabolically labeled with [³⁵S]-methionine and proteins immunoprecipitated with the G-H2CT17 antibody. Lane 1: CVN-transfected NIH 3T3 vector control line. Lanes 2 and 3: Parental and amplified HER2-3 line. Lanes 4, 5, and 6, 7: Parent and amplified lines derived from two independent clones, A1 and B2, of the ATM mutant. The arrows indicate the positions expected for proteins of apparent molecular mass of 175 and 185 kDa.

Fig. 3. Autophosphorylation of p185^{HER2} and p185^{HER2^{ATM}} proteins. Triton X-100 lysates of control, HER2-3₄₀₀, and ATM-expressing cell lines were prepared and immunoprecipitated with the G-H2CT17 antibody. The immune complexes were incubated in 50 ul of HNTG, 5 mM MnCl₂ with 3 uCi [γ -³²P] for 20 min, electrophoresed on a 7.5% polyacrylamide gel, and labeled bands visualized by autoradiography. Lane 1: CVN vector control. Lane 2: HER2-3₄₀₀ cells expressing full-length HER2. Lanes 3 and 4: Two independent lines expressing p185^{HER2^{ATM}}. The arrows indicate the positions expected for proteins of apparent molecular mass of 66.2, 97, 175, and 185 KDa.

Fig 4. Secretion assay of ATM mutants. Cell lines CVN, HER2-3₄₀₀, ATM-A1₄₀₀, and ATM-B2₄₀₀ were labeled with [³⁵S]-methionine overnight. Triton X-100 cell extracts were prepared and the labeling medium collected. Cells and cell-conditioned media were immunoprecipitated with G-H2CT17 antibody and analyzed on 7.5% SDS-PAGE gels.

-7-

Lanes 1-4 are immunoprecipitations of cell extracts from the various lines, and lanes 5-8 are immunoprecipitations from the corresponding cell-conditioned media. Lanes 1 and 5: CVN vector control. Lanes 2 and 6: HER2-3₄₀₀ cell lines expressing full-length p185^{HER2}. Lanes 3, 4 and 7, 8: ΔTM-A1₄₀₀ and ΔTM-B2₄₀₀ cell lines expressing mutant p185^{HER2ΔTM}. The arrows indicate the positions expected for proteins of apparent molecular mass of 175 and 185 KDa.

Fig 5. Secretion of p185^{HER2XCD} from 3T3 and CHO cells. NIH 3T3 and CHO cell lines expressing full-length and truncated p185^{HER2} and vector controls were labeled with [³⁵S]-methionine overnight. Cell extracts and cell-conditioned media were immunoprecipitated with anti-HER2 monoclonal antibody 3E8 and analyzed on 7.5% SDS-PAGE gels. Lanes 1 and 2: NIH 3T3 control cell line, extract and conditioned medium. Lanes 3 and 4: NIH 3T3 line A1 expressing p185^{HER2XCD}, cells and medium. Lanes 5 and 6: NIH 3T3 line 3₄₀₀ expressing full-length p185^{HER2}, cells and conditioned medium. Lanes 7 and 8: CHO control line, cell extract and conditioned medium. Lanes 9 and 10: CHO line 2, expressing p185^{HER2XCD}, cells and conditioned medium. Lanes 11 and 12: CHO line HER2₅₀₀, expressing full-length p185^{HER2}, cells and conditioned medium. The arrows indicate the molecular mass of the indicated protein bands.

Fig 6. Increase in expression of p185^{HER2XCD} with amplification. The CHO-derived cell line HER2XCD-2 was selected for growth in 500 nM and then 3000 nM methotrexate. The parent line, the two amplified

-8-

derivatives, and control vector-transfected cells were labeled with [^{35}S]-methionine. Cell extracts and cell-conditioned media were immunoprecipitated with the anti-HER2 monoclonal antibody 3E8 and analyzed on a 7.5% SDS-PAGE gel. Lanes 1 and 2: CVN cell extract and conditioned medium. Lanes 3 and 4: HER2XCD-2, unamplified cells and conditioned medium. Lanes 5 and 6: HER2XCD-2 amplified to resistance to 500 nM methotrexate, cells and conditioned medium. Lanes 7 and 8: HER2XCD-2 amplified to resistance to 3000 nM methotrexate, cells and conditioned medium. For comparative purposes, one-fifth as much sample of the 3000 nM line was loaded compared to the control, 0 nM, and 500 nM lines. The band intensities were quantitated with an LKB2202 laser densitometer. The arrows show the positions of proteins of apparent molecular mass of 88 and 103 KDa.

Fig 7. Biosynthesis of p185^{HER2XCD}. The CHO line HER2XCD2₃₀₀₀ was labeled with [^{35}S]-methionine and cell extracts, and cell-conditioned media prepared. Lanes 1 and 2: Cell extract and cell-conditioned medium. Lanes 3 and 4: The same conditioned medium incubated or mock-incubated with endo H. Lanes 5 and 6: Cell extract and conditioned medium from cells treated with tunicamycin. The arrows show the positions expected for proteins of apparent molecular mass of 73, 88, and 103 KDa.

Fig 8. Morphology of NIH 3T3 cells transfected with HER2 and HER2 Δ TM expression constructs. A and D: Parental and amplified cells from NIH 3T3 cells transfected with

-9-

vector alone. B and E: NIH 3T3 cells expressing p185^{HER2ΔTM} (line A1), parent and amplified derivative selected for resistance to 400 nM methotrexate. C and F: NIH 3T3 cells expressing wild type p185^{HER2} (line 3), parent and amplified
5 derivative selected for resistance to 400 nM methotrexate.

Fig 9. Cell surface and cytoplasmic immunofluorescence staining of control, HER2, and HER2ΔTM lines. The top photos are intact cells labeled with anti-HER2 monoclonal antibody. The bottom photos are the same cell
10 lines treated with 0.15% Triton X-100 detergent before addition of antibody. A and D: Control NIH 3T3 cells transfected with vector only. B and E: Cell line HER2ΔTM-A1₄₀₀, expressing p185HER2ΔTM. C and F: Cell line HER2-3₄₀₀ expressing p185^{HER2}.

15 Fig 10. Fluorescence-activated cell sorter histograms of control, HER2 and HER2ΔTM cells binding anti-p185^{HER2} monoclonal antibody 4D5. Binding by the control antibody, 368, directed against human tissue plasminogen activator, light, broken line. Binding by the anti-HER2 antibody 4D5,
20 dark unbroken line. Panel A: Control NIH 3T3 cells transfected with vector only. Panel B: Cell line HER2-3₄₀₀, expressing p185^{HER2}. Panel C: Cell line HER2 ΔTMA1₄₀₀ expressing p185^{ΔTM}.

Fig 11. Biosynthesis of p185^{HER2} and p185^{HER2ΔTM} proteins.
25 Cell lines HER2-3₄₀₀ and HER2ΔTM-A1₄₀₀ were labeled with [³⁵S]-methionine and p185^{HER2} and p185^{HER2ΔTM} proteins collected by immunoprecipitation and analyzed on a 7.5% SDS-PAGE gel. Lane 1: Vector control. Lane 2: Untreated p185^{HER2ΔTM}. Lanes

-10-

3 and 4: Aliquots of the same cell extract treated or mock-treated with endo H. Lane 5: Nonglycosylated p185^{HER2} from cells treated with tunicamycin. Lane 6: Untreated p185^{HER2}. Lanes 7 and 8: Aliquots of the same cell extract treated or mock-treated with endo H. Lane 9: Nonglycosylated p185^{HER2ΔTM} from cells treated with tunicamycin. The arrows show the positions of proteins of apparent molecular weight of 175 and 185 kDa.

Fig. 12. Purification of the HER2 extracellular domain. Purified HER2 extracellular domain samples were analyzed using PhastSystem SDS-Gel electrophoresis and silver stained protocols as recommended by Pharmacia. SDS polyacrylamide gel (10-15% gradient) electrophoretic analysis was performed according to Pharmacia protocol File No. 110. Silver staining was performed according to Pharmacia protocol File No. 210. Lane 1 contains molecular weight markers (BRL). Lane 2: Chinese Hamster Ovary Cell X concentrate (1 microliter). Lanes 3 and 4: immunoaffinity purified HER2 extracellular domain (1.6 micrograms and 0.16 microgram, respectively). Lanes 5 and 6: immunoaffinity purified HER2 extracellular domain after DEAE chromatography (0.25 micrograms and 0.083 micrograms, respectively). Lanes 7 and 8: HER2 extracellular domain after formulation in PBS (0.32 micrograms and 0.082 micrograms, respectively).

Fig. 13. The predicted amino acid sequence of the HER2 extracellular domain, with the corresponding nucleic acid sequence. The boxed sequences show potential T-cell

-11-

epitopes, using the algorithm developed by Margolit et al.,
J. Immunol. 138:2213-2229(4) (1987).

DETAILED DESCRIPTION

It was initially hypothesized that removal of the
5 transmembrane spanning sequence would yield a protein which
would be secreted from the cell. As previously indicated,
the transmembrane spanning sequence is principally composed
of hydrophobic amino acids, which effectively anchor the
protein in the cell membrane. Removal of this sequence
10 would therefore be expected to permit passage of the
protein through the membrane.

A first construct was accordingly prepared which
deleted exactly in-frame the 22 amino acid transmembrane
spanning sequence of HER2, and 3 amino acids on either side
15 (Figure 1). The construct was prepared as follows:

The central EcoR1 fragment containing the trans-
membrane spanning segment was cloned into the EcoR1 site of
the bacteriophage vector M13 mp18 (Yanisch-Perron et al.,
Gene, 33:103-119 (1985). The noncoding strand was used as
20 template for oligonucleotide-directed mutagenesis. The
construct deleted the transmembrane spanning sequence, and
an additional 3 amino acids before and after.

Residues 651-678 were deleted by priming double
stranded DNA synthesis with a 30 base pair oligonucleotide
25 of sequence 5' CAG AGA GCC AGC CCT CAG CAG AAG ATC CGG 3'.
The double stranded DNA was transformed into SR101 cells
and mutants identified by hybridization to the same
oligonucleotide 5' end labeled by polynucleotide kinase and

-12-

[γ -³²P] ATP (Amersham, 5000 Ci/mmol). An EcoRI fragment containing the deletion was recombined into a plasmid expressing the HER2 cDNA, replacing the wild type sequence.

When expressed in NIH 3T3 cells, this mutant, designated HER2^{ΔTM}, produced a polypeptide, designated p185^{HER2ΔTM}, of apparent molecular weight 175 kD (Figure 2, lanes 5 and 7). Production took place at levels comparable to wild type p185^{HER2} amplified to the same level of resistance to methotrexate (Figure 2, lane 3). The mutant proteins also retained an active tyrosine kinase activity.

In the presence of [γ -³²P]-ATP, the mutant proteins (Figure 3, lanes 3 and 4) were autophosphorylated to the same extent as unaltered p185^{HER} (Figure 3, lane 2). Figure 3 also shows autophosphorylated p185^{HER2ΔTM}-related proteins of lower molecular weight than the complete protein. These smaller proteins may represent degradation products and, since they are not observed with p185^{HER2}, could imply a difference in intracellular processing of the mutant form.

To determine whether the form lacking the trans-membrane sequence was secreted, cells were metabolically labeled with ³⁵S-methionine. The culture conditions used herein were as follows: cells were cultured in a 1:1 mixture of Dulbecco's modified Eagle's medium and Ham's nutrient mixture F-12 supplemented with glutamine (2 mM), penicillin (100 units/ml), streptomycin (100 ug/ml), and 10% serum. NIH 3T3-derived cell lines were cultured with calf serum (Hyclone). Chinese Hamster Ovary cells deficient in dihydrofolate reductase (CHO-DHFR) were

-13-

cultured in fetal bovine serum (Gibco) supplemented with glycine (0.13 mM), hypoxanthine (0.11 mM), and thymidine (0.02 mM). (For selection of the transfected plasmid DHFR gene or to amplify introduced plasmids by methotrexate selection, the glycine, hypoxanthine, and thymidine were
5 omitted and extensively dialyzed serum substituted for fetal bovine serum.)

Both cells and cell-conditioned medium were assayed for p185^{HER2}. Figure 4 demonstrates that all p185^{HER2} remained
10 cell associated (lanes 2, 3, 4), and neither the wild type protein nor the mutant form was secreted (lanes 6, 7, 8).

Thus, contrary to expectations, deletion of the transmembrane spanning sequence was not sufficient to yield a secreted form of p185^{HER2}.

15 The discovery that p185^{HER2}^{ΔTM} is not secreted suggested that perhaps there are sequences distal to the transmembrane spanning region that prevent passage of p185^{HER2} through the plasma membrane. A second mutant was accordingly made that contained a UAA stop codon 8 amino
20 acids before the beginning of the proposed transmembrane spanning sequence (Figure 1).

The second construct truncated p185^{HER2} 8 amino acids before the start of the transmembrane spanning region at residue 645 by addition of a polypeptide chain-terminating
25 TAA codon. The oligonucleotide 5' AAG GGC TGC CCC GCC GAG TAA TGA TCA CAG AGA GCC AGC CCT 3' was used to prime synthesis of double-stranded DNA from the same template used to construct the ΔTM mutant. Mutant plaques were

-14-

identified by hybridization to the 5' end-labeled oligonucleotide, and confirmed by checking for the presence of a Bcl 1 site also introduced directly after the ochre codon. The chain-terminated mutant, designated HER2^{XCD}, was then recombined into the HER2 cDNA expression plasmid. The structure of the plasmid and the 2 mutant HER2 derivatives is shown in Figure 1.

Secretion of the resulting form of p185^{HER2}, designated p185^{HER2XCD}, was assayed by first metabolically labeling the cells with ³⁵S-methionine, followed by immunoprecipitation of p185^{HER2}-related proteins from both the cells and cell-conditioned media. In the immunoprecipitation procedure (Hudziak et al., Proc. Natl. Acad. Sci. USA, 84:7159-7163 (1987)), cells were harvested by trypsinization, counted electronically with a Coulter counter, and plated at least 6 hrs. before labeling. The plating medium was removed, cells washed with PBS, and the cells re-fed with methionine-free Dulbecco's modified minimal medium. [³⁵S]-methionine (Amersham, 800 Ci/mmol, 29.6 TBq/mmol) was added at 100 uCi/6 cm plate in a volume of 3 ml. Cells were lysed at 4°C with 0.4 ml of HNEG lysis buffer per 6 cm plate. After 10 min, 0.8 ml of lysis dilution buffer (HNEG buffer with 1% bovine serum albumin, 0.1% Triton X-100 detergent) was added to each plate and the extracts were clarified by microcentrifugation for 5 min. Medium to be assayed for secretion of p185^{HER2} related proteins was collected and clarified by microcentrifugation.

-15-

Antibodies were added to cell extracts or conditioned medium and allowed to bind at 4°C for 2-4 h. The polyclonal antibody, G-H2CT17(0), recognizing the carboxy-terminal 17 amino acids of p185^{HER2}, was used for characterization of cell lines expressing the transmembrane-deleted form of p185^{HER2}. The monoclonal antibody 3E8, recognizing an epitope on the extracellular domain (Hudziak et al., Mol. Cell. Bio., 9:1165-1172 (1989)), was used at 8 ug/reaction to immunoprecipitate the truncated form. Seven ug of rabbit anti-mouse IgG was added to immunoprecipitations using this monoclonal to improve its binding to protein A-sepharose. Immune complexes were collected by absorption to protein A-sepharose beads and washed (Hudziak et al., Proc. Natl. Acad. Sci. USA, 85:5102-5106 (1988); Hudziak et al., Proc. Natl. Acad. Sci. USA, 84:7159-7163 (1987)). Proteins were separated on 7.5% sodium dodecyl sulphate-polyacrylamide gels (SDS-PAGE) and analyzed by autoradiography.

This revealed a form of p185^{HER2XCD} of M_r 88,000 kD that is associated with the cells (Figure 5, lanes 3 and 9); however, the cell-conditioned media from both the NIH 3T3 cells and Chinese hamster ovary-derived lines also contains larger amounts of a protein of M_r 103,000, which is immunoprecipitated by anti-HER2 monoclonal antibody (Figure 5, lanes 4 and 10). Full length p185^{HER2} was also expressed in both NIH 3T3 and CHO cells (Figure 5), lanes 5 and 11. There is no secretion of native p185^{HER2} from either of these cell types (Figure 5, lanes 6 and 12).

-16-

The larger size of the observed proteins in the cells and cell-conditioned medium (88,000 and 103,000, respectively) compared to the size predicted by the amino acid sequence (71,644) suggested that the truncated form
5 was being glycosylated.

This was confirmed by treating the cells with the antibiotic tunicamycin, which prevents N-linked glycosylation. Treatment with tunicamycin resulted in the appearance of a cell-associated protein of M_r 73,000, which
10 is close to that predicted by the amino acid sequence (Figure 7, lane 5). It also almost completely inhibited secretion of p185^{HER2XCD} into the medium (Figure 7, lane 6). Cell-conditioned medium from tunicamycin treated cells contains only small amounts of the mature 103,000 form, and
15 none of the smaller forms (lane 6). This further suggests that secretion of p185^{HER2XCD} is coupled to glycosylation.

The extent of glycosylation of the secreted form was investigated with the enzyme endoglycanase H (endo H, Boehringer Mannheim). This enzyme will hydrolyze
20 asparagine-linked oligosaccharides of the high mannose type. High mannose oligosaccharides are biosynthetic intermediates in the glycosylation process. Final maturation of the carbohydrate side chains involves trimming off some mannose and addition of other sugars such
25 as fucose. Such mature oligosaccharide side chains are resistant to endo H.

To determine if secreted p185^{HER2XCD} is resistant to this enzyme, cell conditioned medium labeled with ³⁵S-methionine

-17-

was immunoprecipitated. The immuno-precipitates were collected onto protein A sepharose beads and incubated with endo H. Neither mock incubated (lane 3) nor endo H-treated p185^{HER2XCD} (lane 4) showed any decrease in mobility
5 associated with hydrolysis of the glycosyl side chains, demonstrating that the glycosylation is complete.

Without being bound by any particular theory, these results taken together suggest that the cell-associated form of p185^{HER2XCD} is an intermediate, and that fully mature
10 glycosylated p185^{HER2} extracellular domain is being synthesized and secreted. The lack of secretion of the p185^{HER2ΔTM} protein could be hypothesized to result from the presence of processing information in the transmembrane spanning sequence which is necessary for Golgi transport
15 and targeting of the plasma membrane; however, from these studies it appears instead that transport of tyrosine kinase receptor (or receptor-like) extracellular domain to the cell surface is coupled to proper glycosylation.

Therefore, insertion of the UAA stop codon 8 amino
20 acids before the beginning of the proposed transmembrane spanning sequence yields a fully mature glycosylated p185^{HER2} extracellular domain which is freely secreted by the cell.

Having succeeded in producing a secreted form of p185^{HER2}, the next stage involved investigating whether the
25 amount of secreted protein could be increased by gene amplification. Using the CHO-derived cell line, it was found that the amount of extracellular domain could be increased by methotrexate selection. The amount of

-18-

secreted product increased 29-fold in cells selected for resistance to 500 nm methotrexate, and a further 4.4-fold by selection for resistance to 3000 nm methotrexate (Fig. 6).

5 Thus, a total increase of 128-fold in secreted p185^{HER2XCD} was obtained when this cell line was amplified to resistance to 3000 nm methotrexate, making the production of relatively large quantities of p185^{HER2XCD} possible.

10 To determine whether overexpression of p185^{HER2ATM} results in cell transformation, DNA was introduced in mammalian cells by the CaHPO₄ coprecipitation method (Graham et al., Virology, 52:456-467 (1973)). Five ug of plasmid DNA was added to half-confluent plates of cells (6.0 cm) in 1 ml for 4-6 h. The DNA was removed and the cells shocked 15 with 20% (vol/vol) glycerol. After 2 days for phenotypic expression the selective agent geneticin was added at 400 ug/ml. Clones were picked using glass cloning cylinders with petroleum jelly for the bottom seal. The introduced plasmids were amplified by the methotrexate selection 20 procedure (Kaufman et al., J. Mol. Biol., 159:601-621 (1982)).

When the ATM mutant was expressed in NIH 3T3 cells, primary unamplified colonies after selection had the normal flat nontransformed phenotype (Figure 8, compare photo B 25 with vector control alone, photo A). After the expression level was increased by methotrexate selection, the cells took on the refractile, spindle-shaped appearance of transformed cells and also grew piled up in irregular

-19-

clumps (photo E). This observation is similar to our earlier findings with the unaltered HER2 cDNA (photos C and F, parent and amplified derivatives respectively), and suggests that high levels of expression of the mutant ATM
5 protein were also transforming.

The morphological changes seen at equivalent levels of amplification (400 nm methotrexate) are not as marked for the mutant, implying that the transforming potential of this form of p185^{HER2} may be less. At higher levels of
10 resistance to methotrexate, the ATM cells become even more transformed in appearance.

The plasmid was also negative in a focus-forming assay whereas the wild type HER2 plasmid was positive, further indicating that the transforming potential of p185^{HER2ΔTM}
15 protein is lower. Cells expressing high levels also displayed another property of the transformed phenotype, growth in soft agar. Colony formation in soft agar was determined by harvesting each line to be assayed with trypsin, counting the cells (Coulter counter), and plating
20 80,000 cells per 6-cm dish. The top layer consisted of 4 ml of 0.25% agar (Difco, "purified") over a bottom layer of 5 ml of 0.5% agar. Colonies were counted after 3-4 weeks. Cells from 2 independent clones plated in soft agar gave rise to soft agar colonies with an efficiency comparable to
25 cells expressing the wild type HER2 gene:

-20-

Table I
Soft Agar Colony Formation

	<u>Cell Line</u>	<u># of Soft Agar Colonies</u>
	CVN	0
5	CVN ₄₀₀	0
	HER2-3 ₀	5 +/- 1
	HER2-3 ₄₀₀	208 +/- 27
	ΔTM-A1 ₀	0
	ΔTM-A1 ₄₀₀	205 +/- 62
10	ΔTM-B2 ₀	0
	ΔTM-B2 ₄₀₀	205 +/- 13

Two control lines were used; NIH 3T3 cells transfected with a plasmid expressing only the neo and DHFR genes, and the same line amplified to resistance to 400 nM methotrexate. The number of soft agar colonies arising was determined for both parental and amplified lines of clones expressing either p185^{HER2} or p185^{HER2ΔTM} proteins. Each cell line was plated in triplicate and the value averaged.

Therefore, according to the present invention it has been determined that removal of only the transmembrane spanning sequence does not lead to secretion of p185^{HER2}, unless the entire tyrosine kinase domain is also deleted. Removal of this domain results in proper glycosylation and secretion of the extracellular domain.

In order to obtain purified HER2 extracellular domain working material, Chinese Hamster Ovary Cell Harvest Fluid (CFF) containing recombinant HER2 ECD may be first concentrated by ultrafiltration, and then purified by immunoaffinity chromatography using a HER2 specific MAb

-21-

coupled to CNBr activated Sepharose; other suitable immobilization supports may be used. Concentrated CCF is applied to the affinity column after filtration through a 0.2 micron Millipor filter. Purification cycles are performed as necessary until the desired amount of CCF is processed.

During each cycle of purification, the concentrated CCF is applied and the affinity column is washed to baseline with 0.5 M Tris buffer containing 0.15 M NaCl at a pH of approximately 7.5 (TB). HER2 extracellular domain is then eluted from the column with 0.1 M sodium citrate buffer containing 0.5 M NaCl at a pH of approximately 3.5. The affinity column eluant fractions containing HER2 ECD are pooled and neutralized. The immunoaffinity column is reequilibrated between each purification cycle with TB.

In a second step, the affinity column eluant is buffer exchanged into 25 ml of Tris buffer, at a pH of approximately 7.0 (TB2). The HER2 extracellular domain is then applied to a DEAE Sepharose Fast Flow column, and washed with TB2. The HER2 ECD is removed from the column by step or gradient salt elution in TB2 (containing up to 200 mM NaCl).

After DEAE chromatography, purified HER2 ECD fractions are pooled, exchanged into phosphate-buffered saline, and stored at 2-8° C. The resulting material is substantially pure, i.e., about 99% pure (see Fig. 12).

By means of the present invention it is accordingly possible to produce a secreted, glycosylated p185^{HER2}

-22-

extracellular domain. This opens several possibilities for further research, as well as a broad range of potential therapeutic applications.

As previously stated, the HER2 gene is of particular
5 interest because its amplification has been correlated with certain types of cancer. In a survey of 189 primary mammary gland adenocarcinomas, it was found that 30% contained amplifications of the HER2 gene. Slamon et al., "Human Breast Cancer: Correlation of Relapse and Survival
10 with Amplification of the HER-2/neu Oncogene," Science 235, 177-182 (1987). Amplification was correlated with a negative prognosis and high probability of relapse.

This suggests that of the 120,000 women diagnosed with breast cancer each year, 36,000 will have HER2
15 amplification. Approximately half of these women, or about 15,000, may be expected to exhibit greater than 5-fold amplification, corresponding to nearly half of the 40,000 breast cancer-related deaths each year.

It has been demonstrated that a monoclonal antibody
20 directed against the p185^{HER2} extracellular domain specifically inhibits growth of breast tumor-derived cell lines overexpressing the HER2 gene product; prevents HER2-transformed NIH 3T3 cells from forming colonies in soft agar; and reduces the resistance to the cytotoxic effect of
25 tumor necrosis factor alpha which accompanies HER2 overexpression. Hudziak et al., "p185^{HER2} Monoclonal Antibody has Antiproliferative Effects In Vitro and Sensitizes Human Breast Tumor Cells to Tumor Necrosis

-23-

Factor", Mol. Cell. Biol. 9:1165-1172 (1989). See also,
Drebin et al., "Inhibition of Tumor Growth by a Monoclonal
Antibody Reactive with an Oncogene-Encoded Tumor Antigen",
Proc. Natl. Acad. Sci. USA 83, 9129-9133 (1986) (in vivo
5 treatment with anti-p185 monoclonal antibody asserted to
inhibit tumorigenic growth of neu-transformed NIH 3T3 cells
implanted in mice).

This effect presents the possibility that conditions
characterized by amplification of the HER2 gene may be
10 subject to treatment via Active Specific Immunotherapy.
This therapeutic modality contemplates provoking an immune
response in a patient by vaccination with an immunogenic
form of the extracellular domain. The extracellular domain
(or a derivative thereof, as discussed below) may be
15 combined with a local adjuvant which is safe and effective
in humans, such as alum, Bacillus calmette-Guerin (BCG),
adjuvants derived from BCG cell walls, Detox (Ribi-
immunochem), Syntex-1, or Corynebacterium parvum.
Alternatively, systemic adjuvants, such as Interferon
20 gamma, Interleukin 1, Interleukin 2, or Interleukin 6 may
be suitable. An appropriate dose and schedule would be
selected to maximize humoral and cell-mediated response.

It may also be possible to enhance an immune response
by targeting the immunogen to the immune system, which
25 could lead to more efficient capture of the antigen by
antigen presenting cells, or by directing the immunogen so
that it is presented by MHC Class 1 molecules, since these
usually induce a T-cell response.

-24-

In addition to Active Specific Immunotherapy, it should be possible to use the purified extracellular domain to isolate and characterize the putative ligand. The HER2 ligand may be used in turn to deliver toxin to tumor cells which are overexpressing HER2, such as by molecular fusion of the ligand with toxin, or by chemical cross-linking. Alternatively, patients overexpressing HER2 may be vaccinated with HER2 ligand conjugated to, or in combination with, a suitable adjuvant.

10 A patient overexpressing HER2 will also presumably be overexpressing the HER2 ligand. The ligand-HER2 binding interaction, which is likely to contribute to tumor growth, may be inhibited by blocking free ligand in the patient's serum. This blocking can be accomplished by treating the patient with the HER2 extracellular domain, which will proceed to bind free HER2 ligand, thereby preventing the ligand from binding to the HER2 receptor site.

20 Rather than using the HER2 extracellular domain per se, it may be more desirable to use a derivative which has an increased affinity for the ligand, and/or which has an increased half-life in vivo. Cross-linking on cells is known to improve binding affinity, suggesting that artificial cross-linking can be used to improve the binding ability of the HER2 extracellular domain. The half-life of the extracellular domain in serum can be improved by, for example, fusing the extracellular domain with other molecules present in the serum which are known to have a

-25-

long half-life, such as the Fc-portion of an immunoglobulin molecule.

The present invention has of necessity been discussed herein by reference to certain specific methods and materials. It is to be understood that the discussion of these specific methods and materials in no way constitutes any limitation on the scope of the present invention, which extends to any and all alternative materials and methods suitable for accomplishing the ends of the present invention.

-26-

WHAT WE CLAIM IS:

1. An extracellular portion of the HER2 molecule comprising at least 9 amino acids, essentially free of transmembrane and intracellular portions of said HER2 molecule.
5
2. An extracellular portion of the HER2 molecule comprising an immune epitope, essentially free of transmembrane and intracellular portions of said HER2 molecule.
- 10 3. The extracellular portion as defined by claim 1, in substantially pure form.
4. The extracellular portion as defined by claim 1, having a purity of at least about 99%.
5. The extracellular portion as defined by claim 1,
15 wherein said extracellular portion is antigenic in animals.
6. The extracellular portion as defined by claim 1, further comprising the entire extracellular portion of said HER2 molecule.
7. The extracellular portion as defined by claim 1,
20 conjugated with a peptide having immunogenic properties.
8. The extracellular portion as defined by claim 7, wherein said peptide comprises an immune epitope.
9. Isolated DNA encoding the extracellular portion as defined by claim 1, terminating upstream of the portion
25 encoding the transmembrane domain of said HER2 molecule.
10. The isolated DNA as defined by claim 9, terminating at least 1 base pair upstream of the portion encoding the transmembrane domain of said HER2 molecule.

-27-

11. The isolated DNA as defined by claim 10, terminating about 24 base pairs upstream of the portion encoding the transmembrane domain of said HER2 molecule.

12. The isolated DNA as defined by claim 9, wherein
5 said DNA encodes a sequence of at least 9 amino acids of said extracellular portion, and none of the transmembrane or intracellular portions of said HER2 molecule.

13. An expression vector comprising the isolated DNA as defined by claim 9.

10 14. The expression vector as defined by claim 13, wherein said expression vector is a virus.

15. A cell into which the expression vector as defined by claim 13 has been introduced.

16. The cell as defined by claim 15, wherein said cell
15 is a prokaryote.

17. The cell as defined by claim 15, wherein said cell is a eukaryote.

18. A process for producing an extracellular portion of the HER2 molecule, comprising the steps of:

20 a) ligating the isolated DNA as defined by claim 9 into an expression vector capable of expressing said isolated DNA in a suitable host;

b) transforming said host with said expression vector;

c) culturing said host under conditions suitable for
25 expression of said isolated DNA and production of said extracellular portion; and

d) isolating said extracellular portion from said host.

-28-

19. The process as defined by claim 18, wherein said host cell is a prokaryote.

20. The process as defined by claim 19, wherein said prokaryote is a bacterium.

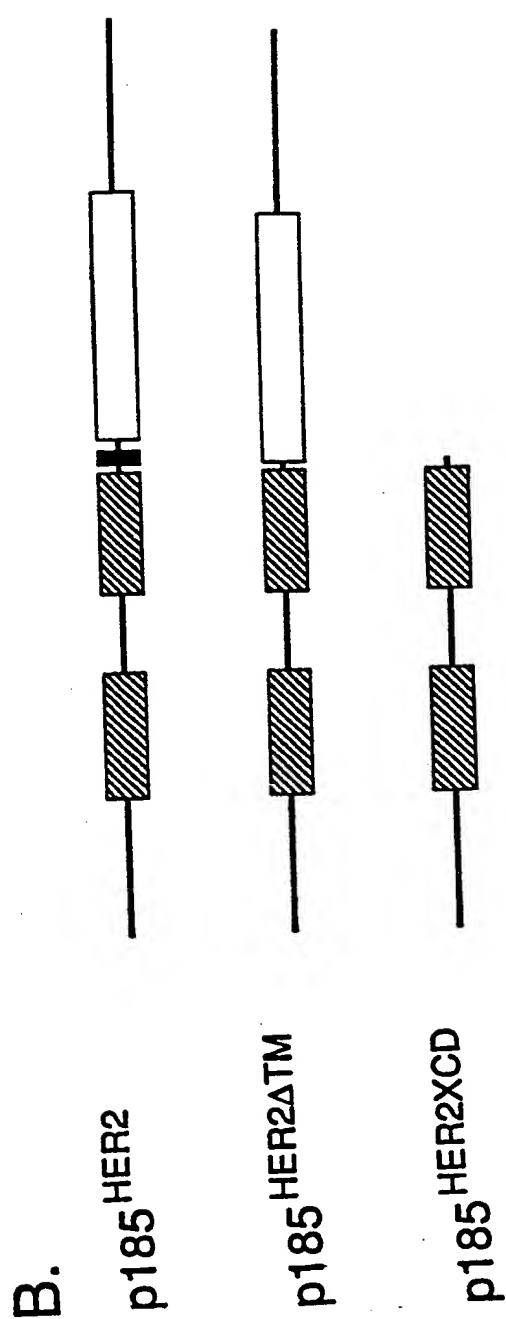
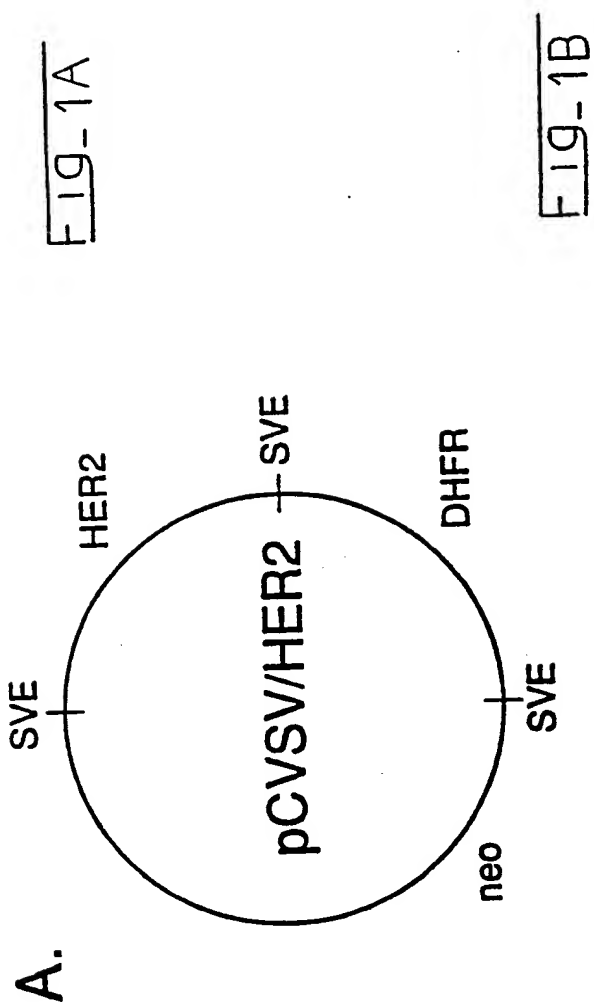
5 21. The process as defined by claim 18, wherein said host cell is a eukaryote.

22. A vaccine comprising the extracellular portion of the HER2 molecule as defined by claim 1.

10 23. The vaccine as defined by claim 22, in combination with a suitable adjuvant.

24. A vaccine comprising the extracellular portion of the HER2 molecule as defined by claim 2.

25. The vaccine as defined by claim 24, in combination with a suitable adjuvant.



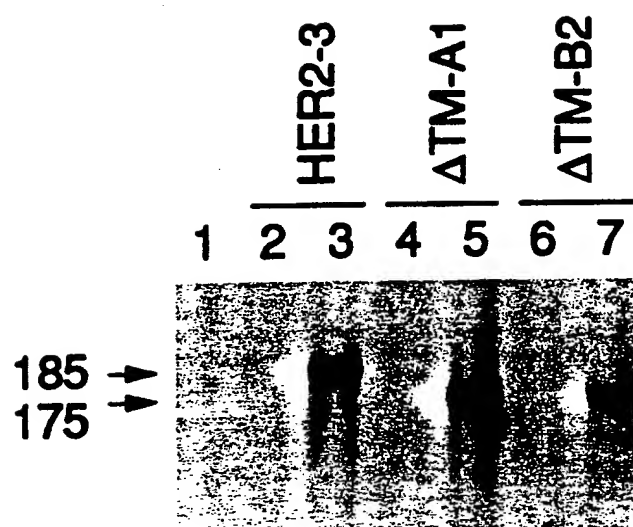


Fig - 2

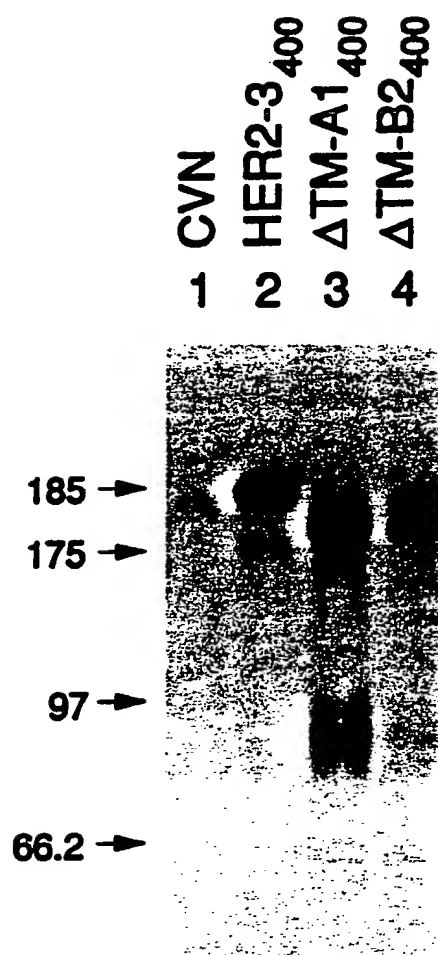


Fig- 3

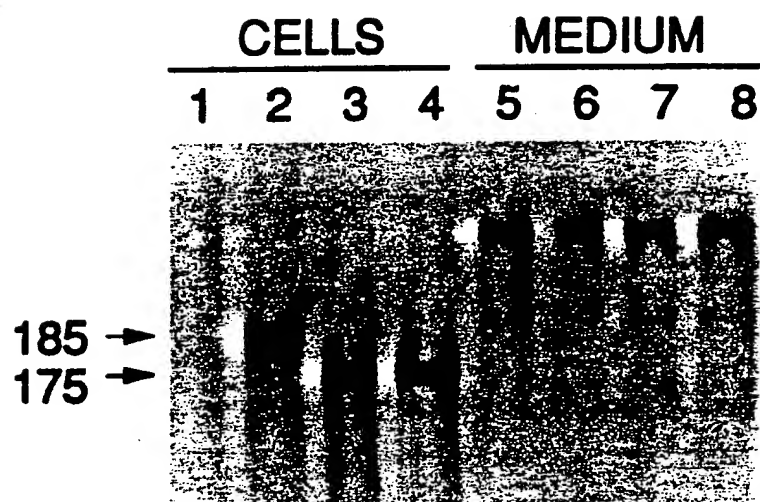
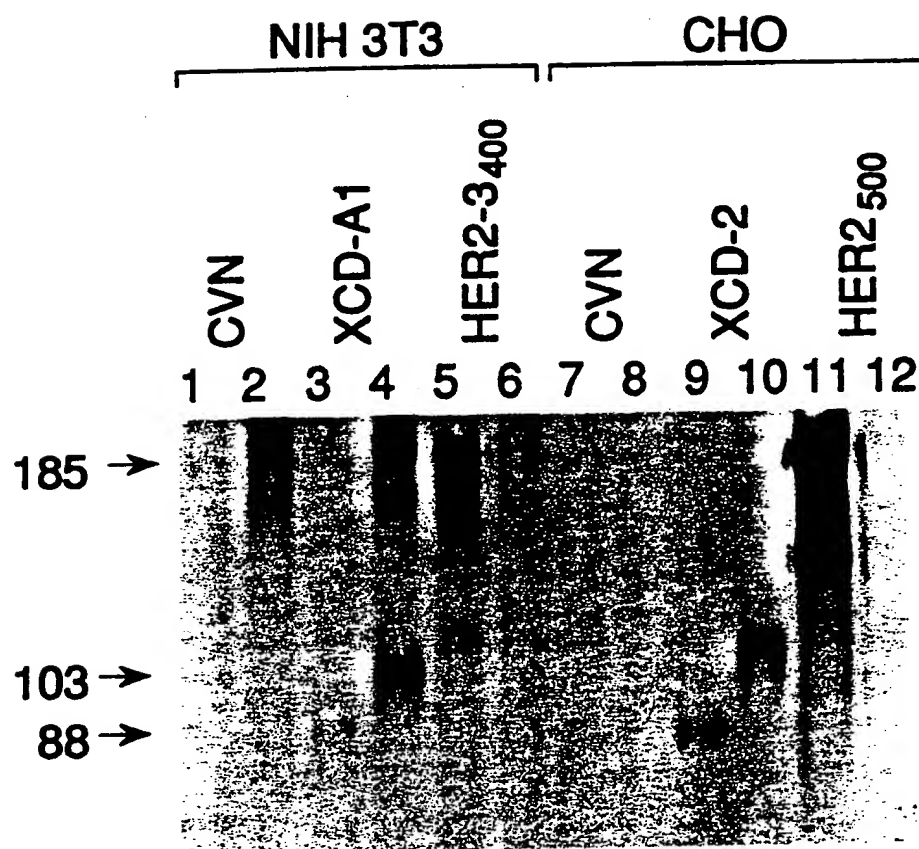


Fig- 4

5/16

Fig - 5

6/16

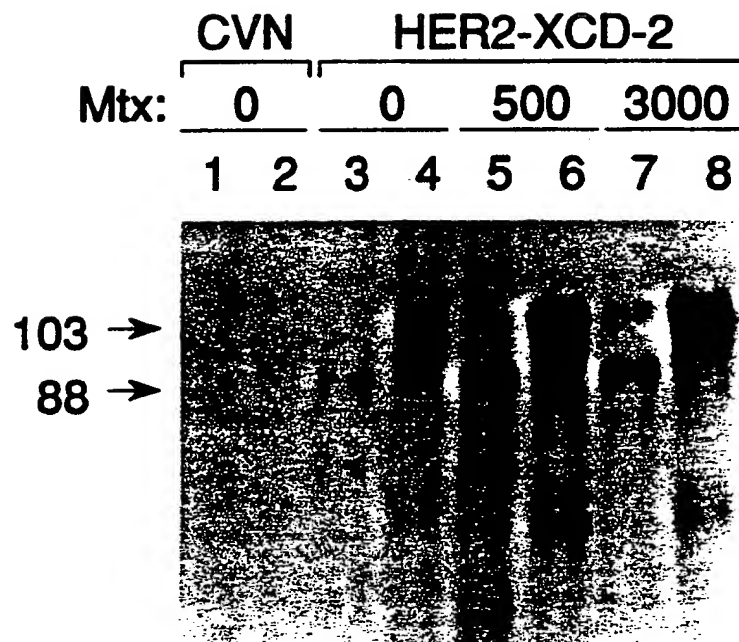


Fig - 6

7/16

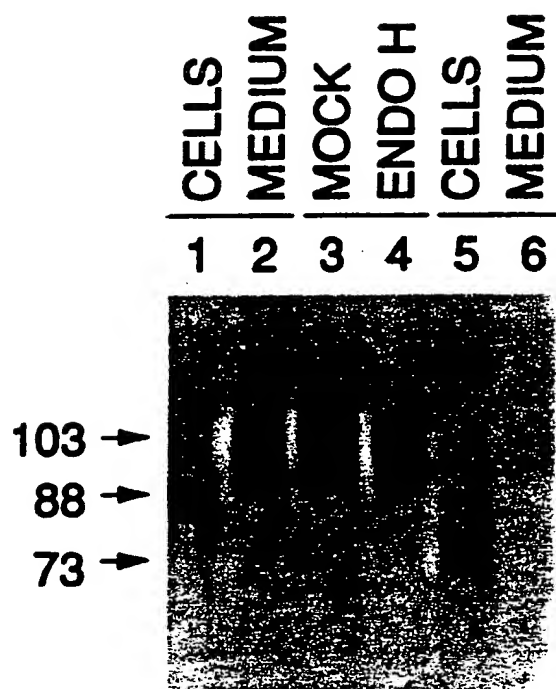
Fig - 7

Fig- 8A

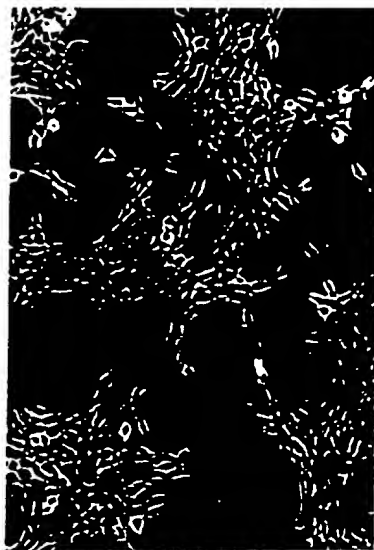


Fig- 8B



Fig- 8C



Fig - 8D



Fig- 8E

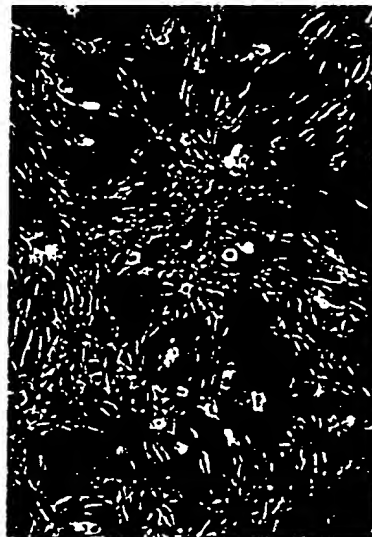


Fig - 8F

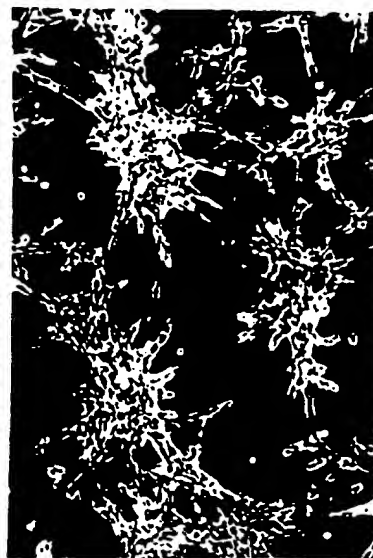


Fig - 9A



Fig- 9B

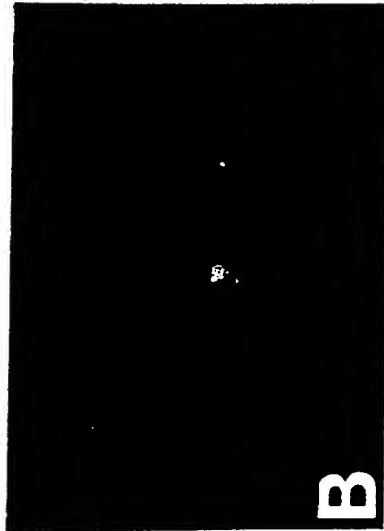


Fig - 9C



Fig- 9D



Fig- 9E

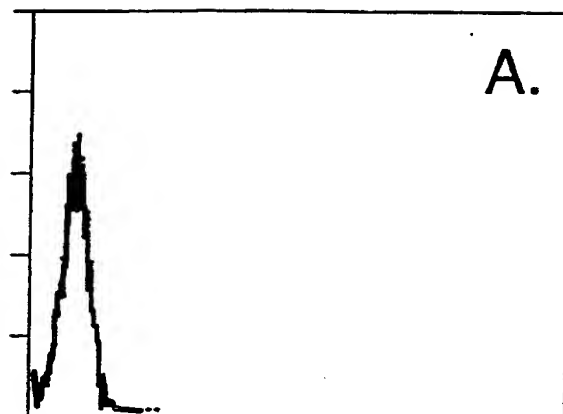
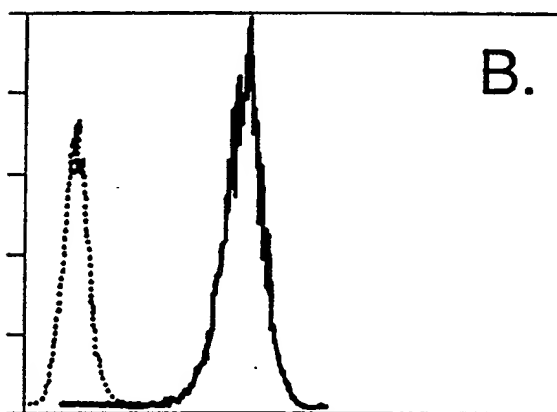
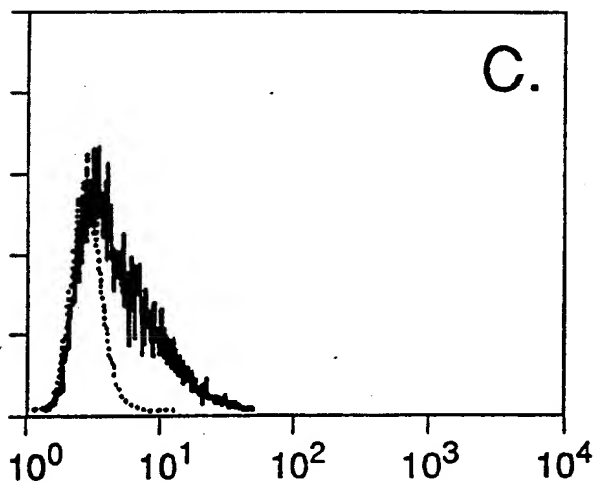


Fig - 9F



10/16

Relative Number of Cells

FIG- 10AFIG- 10BFIG- 10C

Relative Fluorescence Intensity

SUBSTITUTE SHEET

11/16

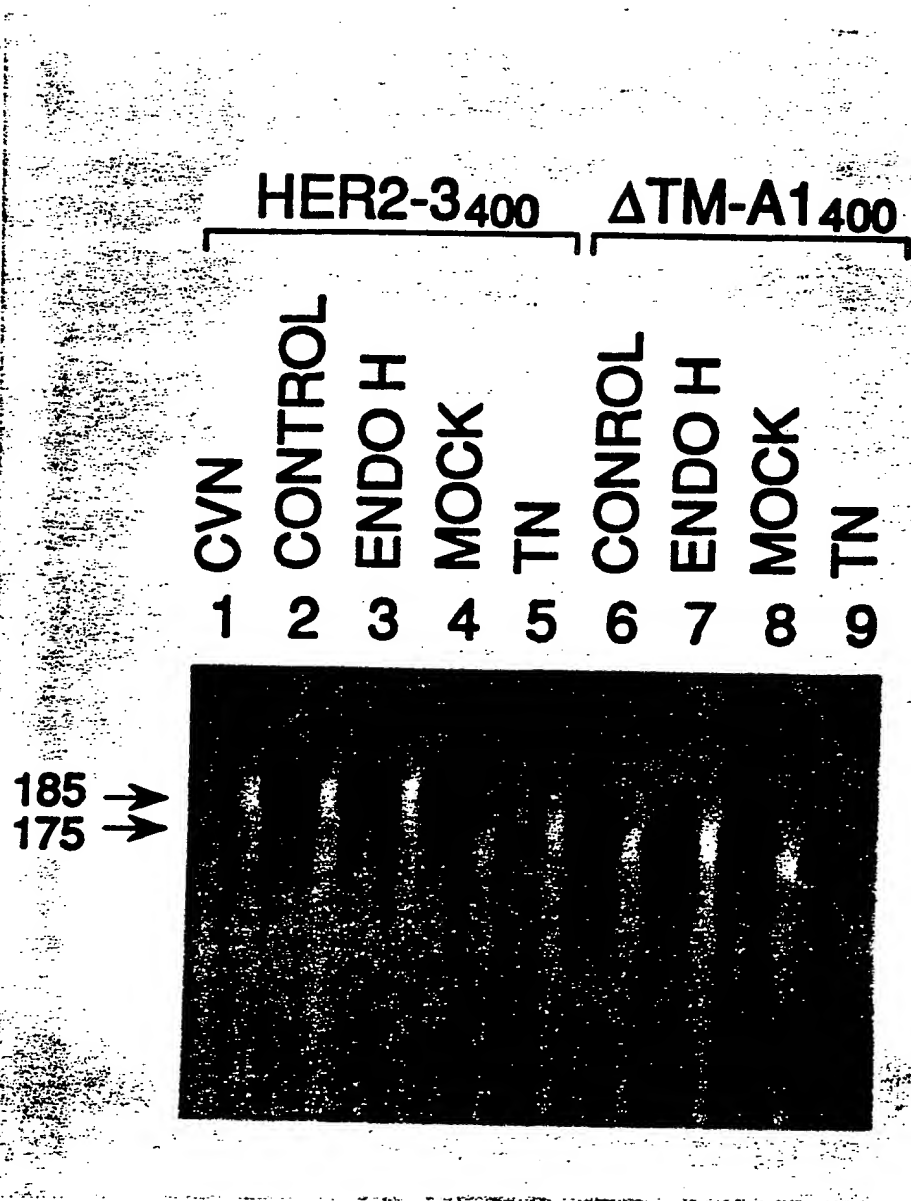


Fig - 11

SUBSTITUTE SHEET

12/16

Purification of the HER2 Extracellular Domain

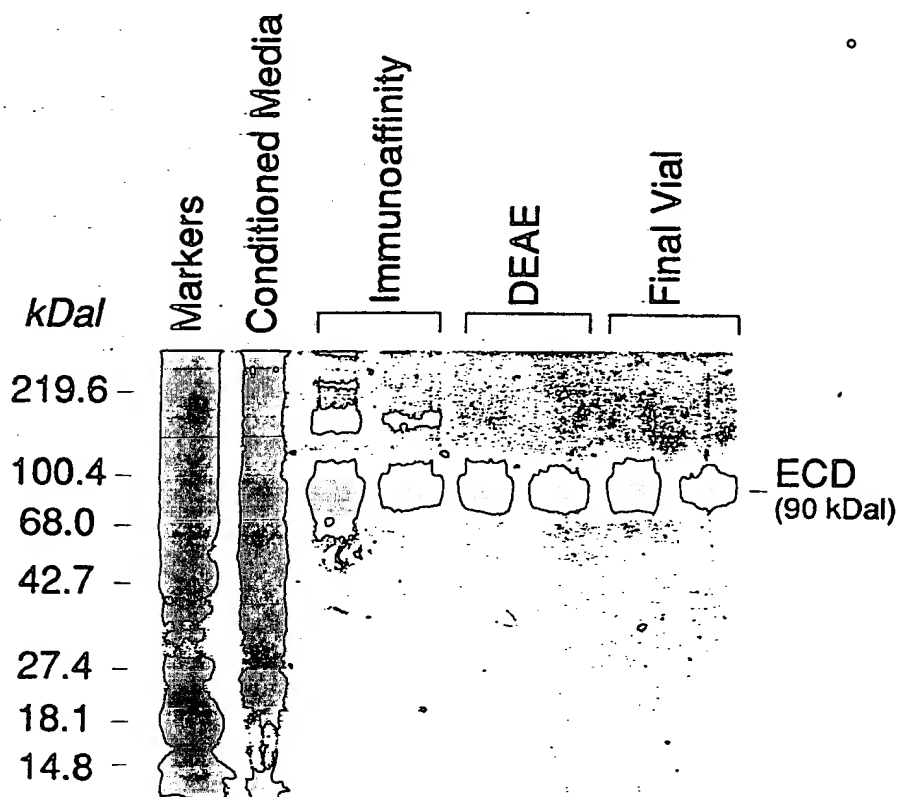


Fig - 12

13/16

FIGURE 13

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1  SER THR GLN VAL CYS THR GLY THR ASP MET LYS LEU ARG LEU PRO ALA SER PRO GLU THR 20
   AGC ACC CAA GUG UGC ACC GGC ACA GAC AUG AAG CUG CGG CUC CCU GCC AGU CCC GAG ACC

   HIS LEU ASP MET LEU ARG HIS LEU TYR GLN GLY CYS 30
   CAC GAC AUG CUC CGC CAC CUC UAC CAG GGC UGC
   GLN VAL VAL GLN GLY ASN LEU GLU 40
   CAG GUG GUG CAG GGA AAC CUG GAA

   LEU THR TYR LEU PRO THR ASN ALA SER LEU SER PHE LEU GLN ASP ILE GLN GLU VAL GLN 60
   CUC ACC UAC CUG CCC ACC AAU GCC AGC CUG UCC UUC CUG CAG GAU AUC CAG GAG GUG CAG

   GLY TYR VAL LEU ILE ALA HIS ASN GLN VAL ARG GLN VAL 80
   GGC UAC GUG CUC AUC GCU CAC AAC CAA GUG AGG CAG GUC
   PRO LEU GLN ARG LEU ARG ILE 80
   CCA CUG CAG AGG CUG CGG AUU

   VAL ARG GLY THR 90
   GUG CGA GGC ACC
   GLN LEU PHE PHE GLU ASP ASN TYR ALA LEU ALA VAL LEU ASP ASN GLY ASP 100
   CAG CUC UUU GAG GAC AAC AAC UAU GCC CUG GUC GUA GAC AAU GGA GAC

   PRO LEU ASN ASN THR THR PRO VAL THR GLY ALA SER 110
   CCG CUG AAC AAU ACC ACC CCU GUC ACA GGG GCC UCC
   PRO GLY GLY LEU ARG GLU LEU GLN 120
   CCA GGA GGC CUG CGG GAG CUG CAG

   LEU ARG SER LEU THR GLU ILE LEU LYS GLY 130
   CUU CGA AGC CUC ACA GAG AUC UUG AAA GGA
   GLY VAL LEU ILE GLN ARG ASN PRO GLN LEU 140
   CUG UUG AUC CAG CGG AAC CCC CAG CUC

   CYS TYR GLN ASP THR ILE LEU TRP LYS ASP ILE PHE HIS LYS ASN ASN GLN LEU ALA LEU 160
   UGC UAC CAG GAC ACG AUU UUG UGG AAG GAC AUC UUC CAC AAC AAC CAG CUG GCU CUC

```


FIGURE 13

350 GLN GLU PHE ALA GLY CYS LYS LYS ILE PHE PHE GLY SER LEU ALA PHE LEU PRO GLU SER PHE
 CAG GAG UUU GCU GGC UGC AAG AAG AUC UUU GGG AGC CUG GCA UUU CUG CCG GAG AGC UUU
 360
 370 ASP GLY ASP PRO ALA SER ASN THR ALA PRO LEU GLN PRO GLU GLN LEU GLN VAL PHE GLU
 GAU GGG GAC CCA GCC UCC AAC ACU GCC CCG CUC CAG CCA GAG CAG CUC CAA GUG UUU GAG
 380
 390 THR LEU GLU GLU ILE THR GLY TYR LEU TYR GGU UAC ACA GGU UAC CUA UAC TYR ILE SER ALA TRP PRO ASP SER LEU PRO ASP
 ACU CUG GAA GAG AUC THR GLY TYR UAC ACA GGU UAC CUA UAC TYR ILE SER ALA TRP PRO ASP SER LEU PRO ASP
 400
 410 LEU SER VAL PHE GLN ASN LEU GLN VAL ILE ARG GLY ARG ILE LEU HIS ASN GLY ALA TYR
 CUC AGC GUC UUC CAG AAC CUG CAA GUA AUC CCG GGA CGA AUU CUG CAC AAU GGC GCC UAC
 420
 430 SER LEU THR LEU GLN GLY LEU GLY GGC AUC AGC TRP LEU GLY LEU ARG SER LEU ARG GLU LEU
 UCG CUG ACC CUG CAA GGG CUG GGC AUC AGC TRP LEU GLY LEU ARG SER LEU ARG GLU LEU
 440
 450 GLY SER GLY LEU ALA LEU ILE HIS HIS ASN THR HIS LEU CYS PHE VAL HIS THR VAL PRO
 GGC AGU GGA CUG GCC CUC AUC CAC CAU AAC ACC CAC CUC CUC UGC UUC GUG CAC ACC GUG CCC
 460
 470 TRP ASP GLN LEU PHE ARG ASN PRO HIS GLN ALA LEU LEU HIS THR ALA ASN ARG PRO GLU
 UGG GAC CAG CUC UUU CCG AAC CCG CAC CAA GCU CUG CUC CAC ACU GCC AAC CGG CCA GAG
 480
 490 ASP GLU CYS VAL GLY GLU GLY LEU ALA CYS HIS GLN LEU CYS ALA ARG GLY HIS CYS TRP
 GAC GAG UGU GUG GGC GAG GGC CUG GCC UGC CAC CAG CAG CUC UGC GCC CGA GGG CAC UGC UGG
 500
 510 GLY PRO GLY PRO THR GLN CYS VAL ASN CYS SER GLN PHE LEU ARG GLY GLN GLU CYS VAL
 GGU CCA GGC CCC ACC CAG UGU GUC AAC AAC CAG AGC CAG UUC CUC UGC CGG GGC CAG GAG UGC GUG
 520

FIGURE 13

530	540
GLU GLU CYS ARG VAL LEU GLN GLY LEU	PRO ARG GLU TYR VAL ASN ALA ARG HIS CYS LEU
GAG GAA UGC CGA GUA CUG CAG GGG CUC	CCC AGG GAG UAU GUG AAU GCC AGG CAC UGU UUG
550	560
PRO CYS HIS PRO GLU CYS GLN PRO GLN GLY LEU	GLY SER VAL THR CYS PHE GLY PRO GLU ALA
CCG UGC CAC CAC CCU GAG CAG UGU CAG CCC CAG AAU	GGC UCA GUG ACC UGU UUU GGA CCG GAG GCU
570	580
ASP GLN CYS VAL ALA CYS ALA HIS TYR LYS	ASP PRO PRO PHE CYS VAL ALA ARG CYS PRO
GAC CAG UGU GUG GCC GCG UGU CAG CAC UAU AAG	GAC CCU CCC UUC UGC GUG GCC CGC UGC CCC
590	600
SER GLY VAL LYS PRO ASP LEU SER TYR MET	PRO ILE TRP LYS PHE PRO ASP GLU GLU GLY
AGC GGU GUG AAA CCU GAC CUC UCC UCC UAC AUG	CCC AUC UGG AAG UUU CCA GAU GAG GAG GGC
610	620
ALA CYS GLN PRO CYS PRO ILE ASN CYS THR	HIS SER CYS VAL ASP LEU ASP ASP LYS GLY
GCA UGC CAG CCU UGC CCC AUC AAC AUC UGC ACC	CAC CAC UCC UGU GUG GAC CUG GAU GAC AAG GGC
624	
CYS PRO ALA GLU	
UCC CCC GCC GAG	

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US90/02697

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ³

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC(5): C07K 7/06, 7/08, 13/00, 17/00; C12N 15/12, 1/19, 1/21 5/10, 15/63
C12P 21/02; A61K 39/00, 39/385, 39/38. U.S.Cl. 530/324-328, 350, 403, 828;

II. FIELDS SEARCHED 435/320, 172, 69.1, 69.3, 252.3, 240.2; 935/12, 22-32, 66-75

Minimum Documentation Searched ⁴ 424/88

Classification System

Classification Symbols

U.S.Cl. 530/324-328, 350, 403, 828; 435/320, 172.3, 69.1, 69.3,
252.3, 240.2; 935/12, 22-32, 66-75; 424/88

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched ⁵

Biosis, World Patents Index, USPTO Automated Patent System (File US PAT,
1975-1990). See attachment for search terms.

III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴

Category ⁶	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁴
X Y	Nature (London, UK), Volume 319, Number 605123, Issued 16 January 1986, Yamamoto et al, "Similarity of Protein Encoded by the Human C- <u>erh</u> -B-2 Gene to Epidermal Growth Factor Receptor", pages 230-234. See Figures 1-3 and final paragraph.	1,2,5,6,9-12 3,4,7,8,13-25
Y	US, A, 4,761,371 (BELL ET AL) 02 August 1988. See the entire document.	7,8,13-21
Y	Journal of Cellular Biochemistry, Supplement 13B, Issued January 1989, Shepard et al, "p185 Her 2 Monoclonal Antibody has Anti-Proliferative Effects <u>in</u> <u>Vitro</u> and Sensitizes Human Breast Tumor Cells to Tumor Necrosis Factor," page 42, abstract D253. See entire document.	3,4,22-25
Y	Biochemistry (Washington, US), Volume 26, number 5, Issued 1987, Yarden et al, "Epidermal Growth Factor Induces Rapid Reversible Aggregation of the Purified Epidermal Growth Factor Receptor", pages 1443-1451. See materials and methods section.	3,4

⁸ Special categories of cited documents: ¹⁵

"A" document defining the general state of the art which is not
considered to be of particular relevance

"E" earlier document but published on or after the international
filing date

"L" document which may throw doubts on priority claim(s) or
which is cited to establish the publication date of another
citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or
other means

"P" document published prior to the international filing date but
later than the priority date claimed

"T" later document published after the international filing date
or priority date and not in conflict with the application but
cited to understand the principle or theory underlying the
invention

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cannot be considered novel or cannot be considered to
involve an inventive step

"Y" document of particular relevance; the claimed invention
cannot be considered to involve an inventive step when the
document is combined with one or more other such docu-
ments, such combination being obvious to a person skilled
in the art.

"A" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search ¹

12 JULY 1990

Date of Mailing of this International Search Report ²

26 SEP 1990

International Searching Authority ¹

RO/US

Signature of Authorized Officer ²⁰

M. Mosher

M. Mosher

Nguyen Ngoc Ho
NGUYEN NGOC HO

INTERNATIONAL DIVISION

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No ¹⁸
Y	Biological Abstracts (Philadelphia, US), Volume 87, Number 10, Issued 15 May 1989, Hudziak et al, "p185 Her 2 Nonoclonal Antibody has Antiproliferative Effects In-Vitro and Sensitizes Human Breast Tumor Cells to Tumor Necrosis Factor", abstract 105430. See entire document.	3,4,22-25
A	Annual Review of Biochemistry (Palo Alto, US), Volume 57, Issued 1988, Yarden et al, "Growth Factor Receptor Kinases", pages 443-78.	1-25

ATTACHMENT TO PCT/US90/02697

SEARCH TERMS

her2, her 2, Ng1, oncogen?, receptor, erb#, erb b, vaccine, pure, purif?,
epidermal growth factor, egf, extra cell?, eternal, ligand, domain,
domains, sequence, C erb B 2, erbb2